

**Date** 2/13/2013

**Objective** PCR amplification of triplet repeat region of FMR1 with a modified program (as below):

- Without hot start
- Three step amplification process
- Denat temp set to 92degC/ 30secs
- Annealing temp set to 62.6deg/ 1:30mins
- Extension temp. grad (55-75deg); time set to 2:30mins

**Description** *Template used:* 100ng genomic DNA NA20230, (53/54 rpt sample from Coriell)  
*Primers:* FT for and FT rev  
*Polymerase:* Native Taq Polymerase (Life Technologies)  
*Solvents:* No Solvt rxns  
 Rxns in 0.5M NMP + 2.2M Betaine

*Reaction Composition:*

Component	0.5M NMP + 2.2M Betaine rxns Final Conc/ amt
Taq Buffer	1 X
MgCl <sub>2</sub>	1.5 mM
dNTPs	0.4 mM
FT-For/ SFS-for	0.4 uM
FT-Rev/ SFS-rev	0.4 uM
gDNA NA203230	100 ng
Taq Polymerase	2.5 U
Water	
NMP	0.5 M
Betaine	2.2 M
<b>total</b>	<b>25ul</b>

*Cycling Conditions:*

- 1 92degC/ 30secs
- 2 62.6degC/ 1:30mins
- 3 55-75degC/ 2:30mins
- 4 GOTO 2 40 times

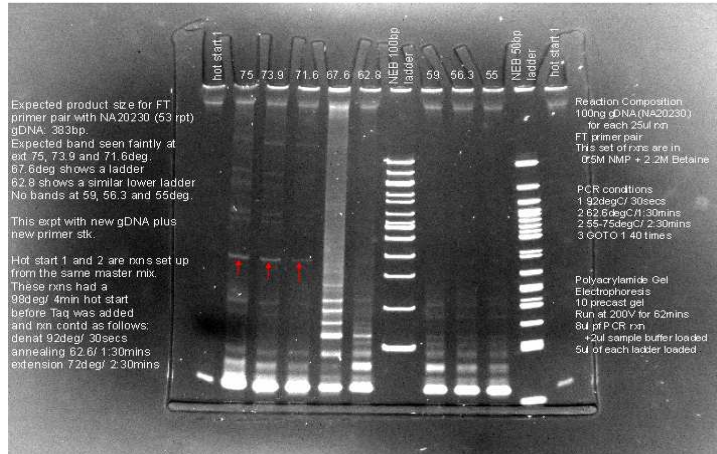
*Gradient steps:* 75, 73.9, 71.6, 67.6, 62.8, 59, 56.3, 55degC

*Gel Electrophoresis:*

10% precast polyacrylamide gels (Life Technologies)  
 8ul of PCR rxn + 2ul of (5X) sample buffer loaded  
 Molecular Ladders: 5ul loaded  
 100bp ladder (NEB)  
 50bp ladder (NEB)

**Gel Pictures**

Lanes are marked according to the ext temp of the particular rxn

**FT primers - NMP + Betaine (20sec exposure)**

**Comments** Expected bands are seen at 75, 73.9 and 71.6deg ext, not very bright (pic is a 20secs exp) Also, other bands are also seen.

The same ext grad (55-75) will also be tried with anng 57.8 to determine which gives better results.